

Multivariate analysis of biochemical responses using non-invasive methods to evaluate the health status of the endangered blackfin goodeid (*Girardinichthys viviparus*)

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ARTICLE INFO

Article history:

Received 28 May 2015

Received in revised form 7 September 2015

Accepted 9 September 2015

Keywords:

Metals

Mucus

PAHs

Alkylphenols

Estrogens

ROS

ABSTRACT

The study of endangered fish species is quite a complex process that involves, in the worst case, the sacrifice of specimens. To solve this ethical problem, some laboratory studies have been conducted in the skin mucus layer (SML) on fish species with valuable results. However, to date research evaluating a panel of biomarkers in the SML of wild fish does not exist. In the current study we assessed the effects of pollutants (E_1 , E_2 , E_3 , EE_2 , BPA, NP, OP, Cu, Fe, Mn, Pb, Zn, $C_{10}H_8$, $C_{16}H_{10}$, $C_{18}H_{12}$, B[a]P, $C_{20}H_{12}$, B[k]F, $C_{20}H_{12}$, $C_{22}H_{12}$) upon a panel of biomarkers [O_2^* , H_2O_2 , lipid peroxidation (as TBARS), carbonyl proteins (RC=O), superoxide dismutase (SOD), catalase (CAT), glutathione peroxidase (GPx), vitellogenin (VTG) and metallothionein (MT)] evaluated in the SML of the wild endangered *Girardinichthys viviparus* inhabiting two polluted lakes in Mexico Valley. Possible relationships were analyzed by principal components analysis (PCA) and redundancy analysis (RDA). The main finding was a clear induction of pro-oxidant forces (ROS) in the SML, probably related to biotransformation of estrogenic and phenolic compounds, in addition to a redox process of Cu and Fe. As a consequence, oxidative tissue damage (TBARS and RC=O) and increases in antioxidant defenses were observed. In male fish, VTG was associated with bisphenol A (BPA) apparently potentiated by Cu and Fe water concentrations; meanwhile, in female fish VTG was linked to estrogens. By PCA, MT was correlated with Fe and Cu; however, it was not linked with diminution of oxidative stress. For the first time have demonstrated the useful of the SML using a panel of biomarkers for monitoring the health of wild fish.

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1. Introduction

Conservation biologists have been increasing the use of novel techniques to study wild vertebrates. In particular, non-invasive approaches for sampling have become the main strategy in wildlife research, allowing the protection of wild specimens (Garshelis, 2006; MacKay et al., 2008). These methods are suitable for protecting endangered species whose existence cannot be further jeopardizing by the use of destructive methods (Esteban and Castaño, 2009; Dzul-Caamal et al., 2013). In the aquatic ecosystem, organisms are exposed to a variety of environmental stressors that generally are present in complex mixtures (Ben-Khedher et al., 2013). During the absorption process, the toxicants released

in aquatic ecosystems are carried by the blood or lymph to a storage point, or to the liver for biotransformation and/or storage, and for excretion through the bile (Needham et al., 2005). However, these substances are likely also translocated to other organs such as the gills or kidney for excretion or could be stored in the fatty tissues (Nussey et al., 2000). As a consequence of these toxicokinetic and toxicodynamic processes, measurements of biomarkers or body burdens on the fluids, cells or tissues detect more the presence of some xenobiotics more quickly and specifically, allowing an early estimation of the damage (Montserrat et al., 2007; Tlili et al., 2010). The skin mucus layer (SML) is the interface between the fish and external environment. It is present on the skin and gill surfaces, but also in the gut lining; indeed, on all mucosal surfaces (Benhamed et al., 2014). The SML plays an important role in immune functions, respiration, ionic and osmotic regulation, reproduction, excretion, and protection against microorganisms, toxicants and hydrolytic enzymes (Cone, 2009; Macpherson et al., 2005).

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In environmental risk assessment, some studies have been conducted in the SML of fish species that found interesting relationships with certain toxicants (Moncaut et al., 2003; Meucci and Arukwe, 2005; Trenzado et al., 2006; Arukwe and Røe, 2008; Rey Vázquez et al., 2009; Maltais et al., 2010; Dzul-Caamal et al., 2013; Nigam et al., 2012, 2014; Maltais and Roy, 2014). However, a wide panel of biomarkers that can be evaluated in the SML of wild fish species has not been available.

On the one hand, Mexico is a privileged land in terms of biodiversity but it is also a country with great vulnerability in its freshwater ecosystems. This phenomenon has historically augmented the degree of disturbance caused by anthropogenic activities, increasing the risk of extinction of native ichthyofauna particularly in Central Mexico (Dzul-Caamal et al., 2012). The blackfin goodeid (*Girardinichthys viviparus* Bustamante, 1839) is a critically endangered fish species according to the IUCN (www.iucnredlist.org) and is included as an endangered species in Mexican guidelines (NOM-059-SEMARNAT-2010). In a previous report, the blackfin goodeid exposed in the laboratory to water from its extant localities suffered a noticeable lipid peroxidation in the liver as well as a variation in the antioxidant defenses according to the sex of the fish were observed (Vega-López et al., 2008a). In the same way, in the liver of the male blackfin goodeid elevated levels of vitellogenin and metallothionein were detected suggesting a greater sensitivity to endocrine disrupting compounds and metals regarding to female fish (Oliveres-Rubio et al., 2015). Interestingly, during the estrogen metabolism carried out by CYP450 isoenzymes ROS induction could be occurring (Cavaliere et al., 2000) and in the goldfish scale cell line some genes involved in the estrogenic response are present such as the osteonectin gene (Lehane et al., 1999). Due to the conservation status of this fish species, the loss and degradation of its habitat and the lack of non-invasive methods for monitoring the health of the blackfin goodeid, the aim of this study was to assess a battery of biomarkers in the SML of *G. viviparus* inhabitants from two polluted lakes in the Valley of Mexico. Biomarkers of the estrogenic response (VTG) and the response to metals (MT) as well as broad-spectrum biomarkers such as pro-oxidant forces ($O_2^{\bullet}$ and H_2O_2), oxidative damage (TBARS and $RC=O$) and antioxidant enzymes (SOD, CAT and GPX) in the SML of *G. viviparus* of both sexes were related to the water concentration of endocrine disrupting compounds (estrogenic and phenolic compounds), heavy metals and polycyclic aromatic hydrocarbons using two multiparametric analyses (principal component analysis and redundancy analysis).

2. Materials and methods

2.1. Sampling and fish collection

Three sampling campaigns were performed in the summer (July), autumn (October) and winter (December) in 2012. Surface and bottom water samples collected with a Van Dorn bottle were placed in 4-L amber glass bottles previously washed and rinsed with HPLC-grade hexane for polyaromatic hydrocarbons (PAH) and estrogen analysis; heavy metals analysis in 1-L plastic bottles.

The collection of specimens was performed in accordance with Mexican protocols for the production, protection and welfare of experimental animals (SAGARPA, 2001); meanwhile, the handling was done in accordance with national and international protocols (<http://eur-lex.europa.eu/LexUriServ/LexUriServ.do?uri=CELEX:31986L0609:EN:NOT>). Thirty *Girardinichthys viviparus* (15♂, 15♀) were collected from each lake using a 3 mm-mesh-opening dip net. The skin mucus layer (SML) was obtained by gently scraping the body surface with a sterile stainless steel spatula into micro-centrifuge tubes. The mucus of the ventral skin was not collected to avoid intestinal contamination and/or intestinal

damage (Dzul-Caamal et al., 2013). After sampling, the fish were placed in situ for observation during 2 h in an 80-L tank filled with aerated water from the lakes before their release. A group of parent fish was maintained in the laboratory for at least one year until reproduction to obtain unexposed fish for use as controls. For maintenance, a glass aquarium of 140 L with synthetic water (0.22 g $MgSO_4$, 0.18 g $NaHCO_3$, 0.08 g KCl and 0.13 g $CaSO_4 \cdot 2H_2O$ per L), photoperiods of 18 h light and 8 dark, at $20 \pm 2^\circ C$, with aeration and constant filtration were used. The eight-month-old control specimens had a mean length of 47.13 ± 0.27 mm and a mean weight of 0.91 ± 0.03 g.

The weight/size ratio or Fulton's condition factor in wild fish ($W = aSL^x$), where W = weight and SL = standard length, were obtained through a potential curve fitting using Microsoft Excel® (Microsoft, Redmond, WA, USA), as previously reported (Le Cren, 1951). The constants " a " and " x " are derived from the mathematical model. The relative condition factor (K) of the fish was calculated as $K = W/aSL^x$ based on Le Cren (1951).

2.2. Sample treatment

The SML was diluted 1:1 (w:v) by the addition of phosphate buffered saline solution (10 mM Na_2HPO_4 , 138 mM NaCl, 2.7 mM KCl, pH 7.4) containing aprotinin (0.04 IU/ml) and 4-(2-aminoethyl) benzenesulphonyl fluoride (2 mM). After homogenization with a hand-held pestle homogenizer the mixture was centrifuged at $9000 \times g/30$ min/ $4^\circ C$. The supernatant (S9 fraction) was stored at $-20^\circ C$ until analysis of antioxidant enzymes and for metallothionein and vitellogenin quantification, except for ROS quantification in which case analysis was performed immediately. A uncentrifuged fraction was re-suspended on a vortex with 1 mL phosphate buffered saline solution for the evaluation of oxidative damage biomarkers, this fraction was chosen because the oxyradicals involved in the initiation of lipid peroxidation in the cellular membranes as well as oxidation and damage to proteins are present in this fraction, not only in the soluble fraction (Hermes-Lima, 2004).

2.3. Quantification of ROS ($O_2^{\bullet}$ and H_2O_2)

The evaluation of ROS ($O_2^{\bullet}$ and H_2O_2) was performed by the specific oxidation of dihydroethidium (DHE) and dihydrofluorescein diacetate (DHF-DA) to determine $O_2^{\bullet}$ and H_2O_2 content, respectively following the Dzul-Caamal et al. (2013) report. Results were expressed per g tissue.

2.4. Lipid peroxidation (TBARS) and protein carbonyl contents ($RC=O$)

Lipid peroxidation evaluated as thiobarbituric acid reactive substances (TBARS) was assessed using the method of Buege and Aust (1978) and results were presented as μ mol TBARS/g tissue.

The oxidation of proteins ($RC=O$) was evaluated by the method of Levine et al. (1994) with some modifications according to previous report (Dzul-Caamal et al., 2013) expressed as mM $RC=O$ /mg protein/g tissue.

2.5. Antioxidant enzymes

The activities of the enzymes involved in antioxidant defense were evaluated in the S9 fraction of the SML. SOD (EC 1.15.1.1) activity was assayed according to the method of Misra and Fridovich (1972). Catalase (CAT) (EC 1.11.1.6) activity was assessed by the method of Radi et al. (1991). The activity of glutathione peroxidase (GPx) (EC 1.11.1.9) was quantified by the method of Lei et al.

(1995). The results for the antioxidant enzymes were expressed as mM/min/mg protein/g of fish.

All tests were performed in triplicate including ROS, oxidative damage biomarkers and antioxidant enzymes.

2.6. Metallothionein (MT) and vitellogenin (VTG) evaluation by enzyme-linked immunosorbent assay (ELISA)

The contents of metallothionein (MT) and vitellogenin (VTG) in the SML were measured by ELISA following the procedure of Vega-López et al. (2006, 2007a) using polyclonal serum (anti-VTG and anti-MT produced in rabbits in the lab) diluted 1:7000. The VTG was quantified by comparison with a standard curve (0.3–6.0 ng) with a purified VTG from *G. viviparus* as standard (Vega-López et al., 2006). MT induction and purification were performed as in Pedersen et al. (1994) and were quantified by a calibration curve (1–30 ng) of *G. viviparus* MT. Results were expressed as ng/mg⁻¹ protein.

2.7. CYP1A expression in the SML

In an independent sampling campaign (August 2013), we evaluated CYP1A1 mRNA in the SML of some male and female wild fish from the lakes under study by RT-PCR because we found statistical relationships between ROS levels with estrogen concentration in the water. Total RNA was isolated using Trizol method, the cDNA synthesis was performed with reverse transcriptase. The first-strand cDNA was employed as the template for RT-PCR with specific CYP1A1 primers (Nogueira et al., 2010) amplifying a product of 500 pb. As an internal control of RNA integrity and cDNA quality, the 18s rRNA gene was used amplifying a product of 453 pb (Vega-López et al., 2007a).

2.8. Physicochemical water assay

2.8.1. Heavy metal analysis

Concentrations of Cu, Fe, Mn, Pb and Zn were determined in acid digestion (HNO₃ AA grade, Sigma–Aldrich™) followed by A.A. using the direct air-acetylene flame method (APHA et al., 1998). Standard curves for Cu, Fe, Mn and Pb were performed from 1.0 to 5.0 mg/L, and for Zn from 0.2 to 1.5 mg/L.

2.8.2. Analysis of estrogenic and phenolic compounds

Estrogen extraction and method for quantification was performed following the protocol of Wang et al. (2006). Curves were constructed from 1–5 ng/L, using Sigma–Aldrich standards and methanol (HPLC grade) as diluent. Retention times were the following: E₁ = 19.9403; E₂ = 16.8064; E₃ = 18.4182; EE₂ = 19.7961 min. The concentration of estrogens was evaluated by the percentage recovery of spiked samples added to standard solutions containing, 1, 10, 50 ng mL⁻¹ of E₃, E₂, E₁ and EE₂, respectively.

The measurement of total phenol (TP) was performed following spectrophotometric EPA method 420.1 by chloroform extraction. Nonyl phenol (NP) was measured by an Alkylphenol ELISA kit (Ecologiena™; Tokiwa Chemical Industries CO., Ltd) with 100% reactivity with NP and 96% with octyl phenol (OP). Bisphenol A (BPA) was measured with a BPA ELISA kit (Ecologiena™; Japan EnviroChemicals, Ltd). These measurements were performed according to the procedures of the manufacturers.

2.8.3. Analysis of polycyclic aromatic hydrocarbons (PAH)

The analysis of PAHs was conducted in a Biotek Synergy MX spectrofluorometer using a Sulpeco™ standard PAH mixture according to a previous report (Vega-López et al., 2013). PAH

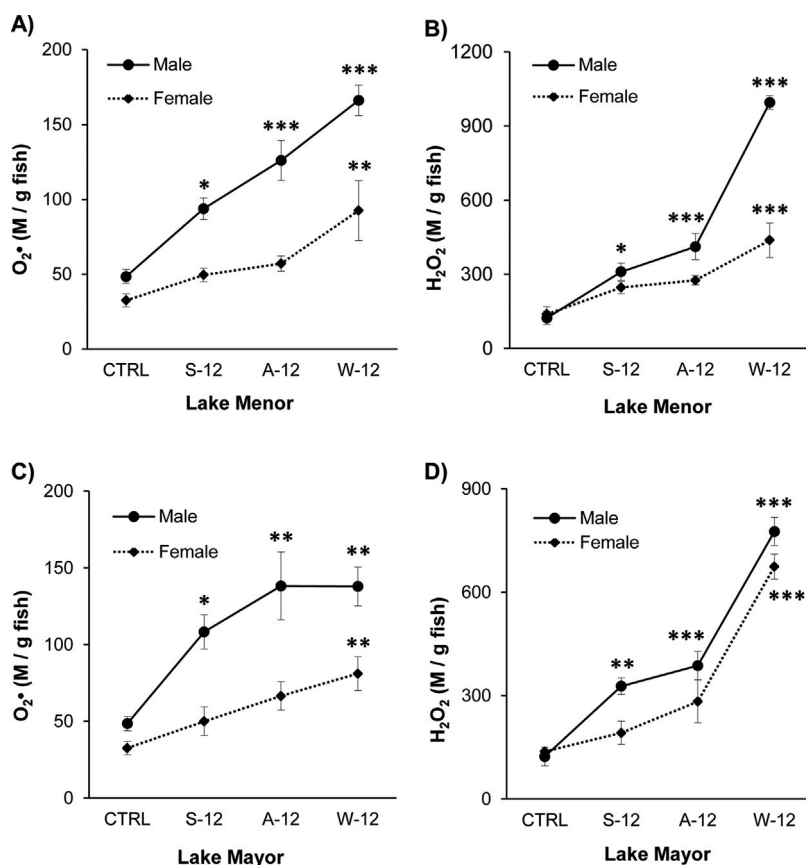


Fig. 1. Content of ROS (O₂* and H₂O₂) in the skin mucus layer (SML) of wild *Girardinichthys viviparus*. Panel (A) and (B) male and female fish from Lake Menor, respectively. Panel (C) and (D) male and female fish from Mayor Lake, respectively. Significant differences with regard to control fish: **p* ≤ 0.05, ***p* ≤ 0.01 and ****p* ≤ 0.001.

Table 1Population features of male and female *Girardinichthys viviparus* from Lake Menor and Lake Mayor of Chapultepec Park (Mexico).

Population features	Menor lake		Mayor lake	
	Males	Females	Males	Females
W (g)	0.35 ± 0.09 ^a	0.70 ± 0.34	0.41 ± 0.08 ^b	0.71 ± 0.30
L-U (g)	0.2–0.56	0.30–1.72	0.25–0.60	0.37–1.87
SL (mm)	26.58 ± 1.64	36.21 ± 4.41	26.58 ± 1.64	34.91 ± 7.76
L-U (mm)	24–31	25.9–48	23.1–30.4	26.7–57.1
K	1.02 ± 0.25 ^a	1.08 ± 0.43 ^a	0.99 ± 0.17 ^b	1.04 ± 0.31 ^b

$n = 45$ fish of each sex and locality. Mean ± standard deviation. W = weight. SL = standard longitude. L = lower value. U = upper value. K = relative condition factor. Different letters denote statistical differences between localities whiting the same sex at $p \leq 0.05$. The relative condition factor (K) of the fish was calculated as $K = W/aSL^x$ using the weight/size ratio or Fulton's condition factor ($W = aSL^x$), where W = weight and SL = standard length based on Le Cren (1951).

standard curve samples (0.002–0.012 $\mu\text{g/L}$) were placed in 96 well plates (polymer base optical button black treated N/SterPS; Nunc) for fluorescence. The excitation and emission wavelengths were as follows: naphthalene (C_{10}H_8) 273 and 360 nm; pyrene ($\text{C}_{16}\text{H}_{10}$) 327 and 385 nm; benzo[a]anthracene ($\text{C}_{18}\text{H}_{12}$) 277 and 376 nm; benzo[a]pyrene ($\text{C}_{20}\text{H}_{12}$) 380 and 430 nm; benzo[k]fluoranthene ($\text{C}_{20}\text{H}_{12}$) 255 and 420 nm; indeno[1,2,3-c,d]pyrene ($\text{C}_{22}\text{H}_{12}$) 250 and 495 nm.

2.9. Statistical analysis

The results were expressed as mean ± standard deviation ($n = 15$, per sex, lake and sampling campaign). Data on the biomarkers were compared using a one-way ANOVA followed by Dunnett post hoc test considering $p \leq 0.05$ to represent a significant difference. In addition, the data was subjected to multiparametric analysis. Principal component analysis (PCA) is based in a linear response model relating species and environmental variables (Van den Brink et al., 2003), which in the current study was used to relate the biomarkers in the SML to environmental variables. PCA results were considered significant with a factorial weight >0.7 . To determine which factors were responsible for the structure or groupings obtained in the PCA, a redundancy analysis (RDA) was carried out. The RDA is an ordination method of direct gradient analysis (Ter Braak and Prentice, 1988) able to find both specific patterns of biological parameters and to assess the relationships between biological (response variables) and environmental data (explanatory variables) using XLSTAT software for Excel. The Monte Carlo permutation was used to evaluate the statistical significance of the canonical axes.

3. Results

3.1. Physiological features of *Girardinichthys viviparus*

The mean weight and standard longitude of wild female fish from both lakes were similar. In contrast, in male fish from Lake Mayor a higher weight with regard to specimens from Lake Menor was documented, whereas the mean standard longitude was similar. However, the specimens of both sexes from Lake Menor presented a higher relative condition factor (K) compared with fish from Lake Mayor (Table 1).

3.2. Content of ROS

The concentration of H_2O_2 was greater than that of O_2^* in the SML of both sexes of wild fish, as observed also in control fish. In the SML of male fish from both lakes (Fig. 1A and C), the concentrations of O_2^* and H_2O_2 were higher in all sampling campaigns than in control fish, with a peak in the winter (December 2012). In the SML of male fish from Lake Menor, maximum levels of O_2^* and H_2O_2 in the SML were 2.8-fold higher and 6.3-fold higher, respectively, than

in the control group. In contrast, in female fish in both lakes, these pro-oxidant forces were significantly greater than in female control fish only in the winter ($p < 0.001$) probably could be explained by high standard deviation observed during the summer and autumn (Fig. 1A–D).

3.3. Oxidative damage biomarkers

In wild fish of both sexes, oxidation of polyunsaturated fatty acids and proteins was greater in all sampling campaigns than in control fish ($p \leq 0.05$). In the SML of male fish, lipid peroxidation evaluated as TBARS increased as ROS concentration increased, particularly in Lake Menor specimens ($p < 0.001$). In females from both lakes, the maximum TBARS level was detected in the autumn ($p < 0.001$) (Fig. 2A). Protein oxidation quantified as carbonyl group (RC=O) content in the SML was higher in fish from both lakes than in control fish, except for male fish from Lake Mayor during the summer. A peak of RC=O was noticeable in the autumn in fish of both sexes (Fig. 2B).

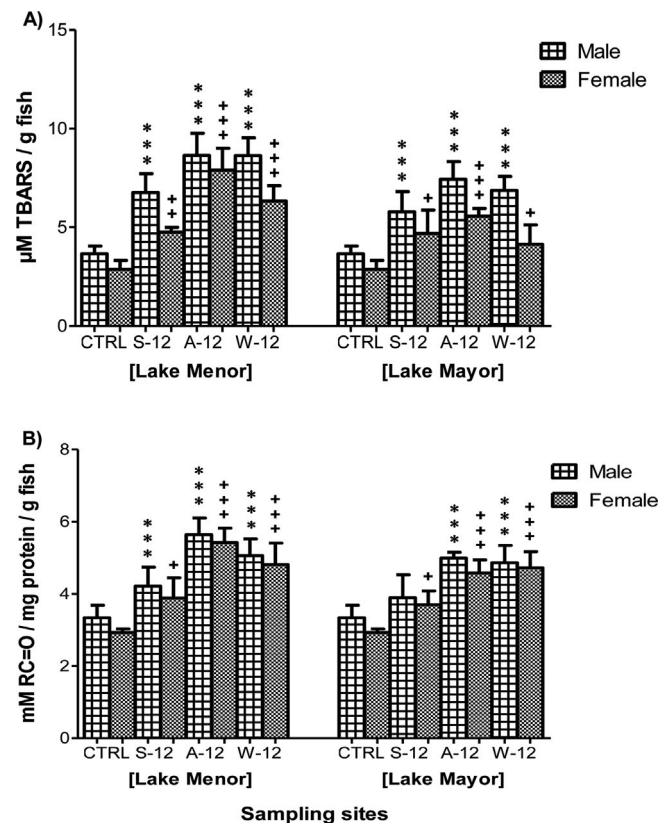
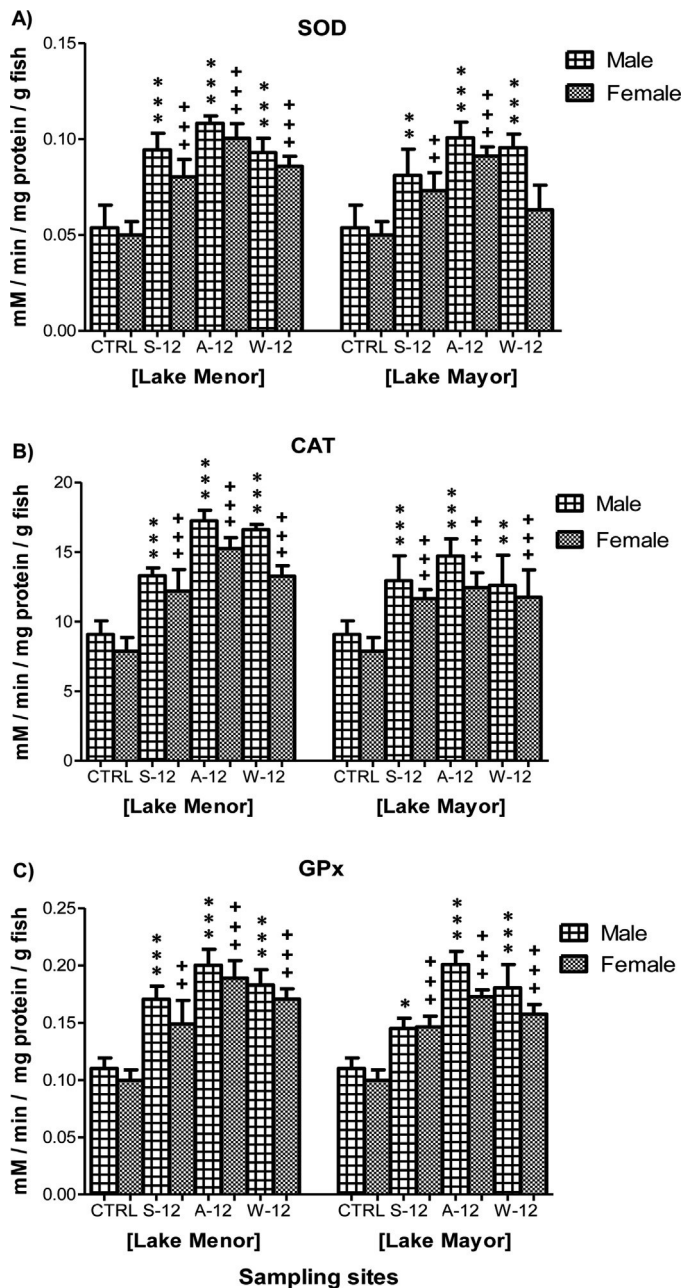


Fig. 2. Levels of lipid peroxidation (A: TBARS) and protein oxidation (B: RC=O) in the skin mucus layer (SML) of wild *Girardinichthys viviparus*. Significant differences with regard to control fish: * $p \leq 0.05$, ** $p \leq 0.01$ and *** $p \leq 0.001$.



3.4. Activity of antioxidant enzymes

In all cases, the activities of the enzymes involved in antioxidant defense in the SML of the wild fish were greater than those in control fish ($p \leq 0.01$). The SOD activity of specimens of both sexes and lakes was elevated statistically ($p \leq 0.01$) compared to control fish except in female fish collected in Lake Mayor during the winter (Fig. 3A). CAT activity was close to two fold higher in wild fish ($p \leq 0.01$) than in control fish. Maximum CAT activity was detected in fish of both sexes from Lake Menor during the autumn, being greater in males than in females (Fig. 3B). GPx showed a higher activity in wild male fish than in females, particularly in the autumn ($p < 0.001$). In addition, the activity of GPx (Fig. 3C) displayed a

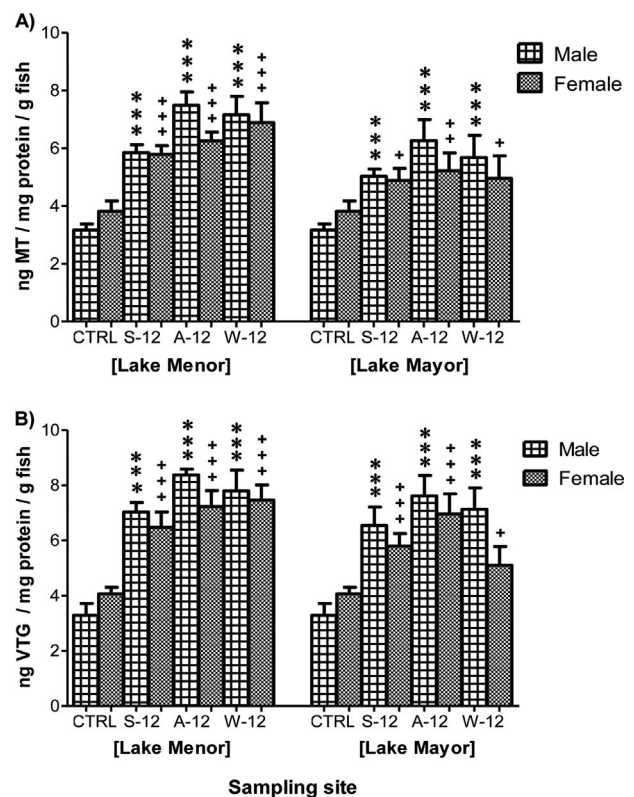


Fig. 4. Levels of metallothioneins (MTs) and vitellogenin (VTG) in the skin mucus layer (SML) of wild *Girardinichthys viviparus*. Panel (A) MT. Panel (B) VTG. Significant differences with regard to control fish: * $p \leq 0.05$, ** $p \leq 0.01$ and *** $p \leq 0.001$.

similar pattern as lipid peroxidation. Interesting, the metabolic rate of GPx was similar to SOD activity, but substantially lower than CAT activity, suggesting the importance of CAT in antioxidant defense in the SML of *G. viviparus*.

3.5. Metallothionein (MT) and vitellogenin (VTG) assays

Levels of metallothionein (MT) in the SML of wild fish of both sexes were greater than in the control group. In male fish from both lakes, a high MT content ($p < 0.001$) with regard to control fish was detected peaking in the autumn, particularly in specimens from Lake Menor. However, in female fish from Lake Menor increased levels of MT reached a peak in the winter ($p < 0.001$); but, in females from Lake Mayor only a slight induction in this biomolecule ($p \leq 0.05$) was found in all sampling campaigns (Fig. 4A).

In wild fish, the amount of vitellogenin (VTG) in the SML was about two-fold higher than in control fish. During the autumn and winter, a noticeable VTG induction in male fish from both lakes and in females from Lake Menor was documented ($p < 0.001$). However, in the winter the amount of VTG in female fish from Lake Mayor decreased with regard to the previous sampling campaigns and was different ($p < 0.05$) from control females (Fig. 4B).

3.6. CYP1A expression

Interestingly, in the SML of male and female fish from Lake Menor and Lake Mayor CYP1A1 expression was observed (SAMEA3420836 ERS739729) (<http://www.ebi.ac.uk/ena/data/view/ERS739729>)

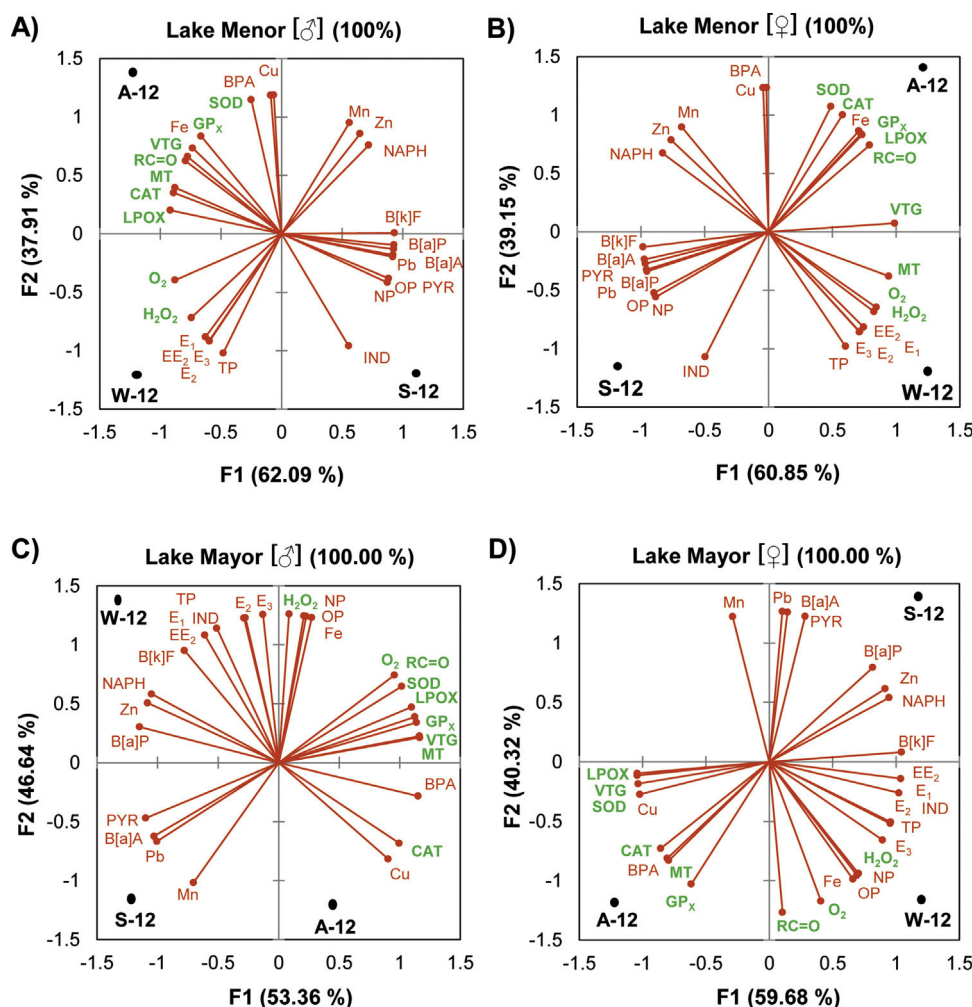


Fig. 5. Principal component analysis (PCA) showing 100% of the explained variance in all cases taking into account the first (F1) and second components (F2) of the panel of biomarkers (O_2^* , H_2O_2 , TBARS, $RC=O$, SOD, CAT, GPx, VTG and MT) evaluated in the SML of wild *Girardinichthys viviparus* and their relations with water levels of pollutants (E_1 , estrone; E_2 , 17 β -estradiol; E_3 , estriol; EE_2 , ethinyl estradiol; TP, total phenols; NP, nonyl phenol; OP, octyl phenol; BPA, bisphenol A; IND, indeno[1,2,3-*c,d*]pyrene; NAPH, naphthalene; PYR, pyrene; B[a]A, benzo[a]anthracene; B[k]F, benzo[k]fluoranthene; B[a]P, benzo[a]pyrene; Fe, Cu, Mn, Pb and Zn). S-12: summer 2012; A-12: autumn 2012; W-12: winter 2012.

3.7. Multivariate analysis of biochemical responses

The obtained results of the principal component analysis (PCA) showed integration of the biomarker responses in the SML of *G. viviparus* between sexes, sites, sampling campaigns and toxicants levels, indicating that discrimination between locations occurs. Taking the first and second components into account explained 100% of the variance in the PCA in all cases (Fig. 5).

Relationships of biomarkers evaluated in the skin mucus layer (SML) were found with phenolic compounds (Fig. 6A), metals (Fig. 6B), estrogens (Fig. 6C), and in a few cases with polyaromatic hydrocarbons (Fig. 6D).

A redundancy analysis plot (RDA) was also constructed with response variables (biomarkers) and explanatory variables (environmental data) (Fig. 7). In fish from Lake Menor the first bi-plot (RDA1) explained a high correlation between the response variables and explanatory variables, both in male (75.01%) and female fish (66.50%). In blackfin goodeid from Lake Mayor the ordination (RDA1) explained 81.28% and 66.50% of the variation in the data in male and female fish, respectively. The triplot (2-dimensional bi-plots; RDA1 and RDA2) explained 100% of the correlation between the response variables and explanatory variables. Also the biomarkers in the SML were related with environmental data.

4. Discussion

The useful of the skin mucus layer (SML) as a target in environmental monitoring represents a significant advance in the protection of endemic or endangered species without requiring their sacrifice. Using multiparametric analysis a sex-linked response in the SML biomarkers of *G. viviparus* with certain environmental pollutants were detected. During the winter, levels of ROS (O_2^* and H_2O_2) were related to water total phenol and to estrogenic compounds (E_1 , E_2 , E_3 and EE_2) in male fish from Lake Menor and in female fish from both lakes using PCA and RDA, suggesting true relationships. In male fish from Lake Mayor, ROS content was dependent on high water levels of certain estrogens (E_2 and E_3) and on the concentration of TP in water using RDA. The present results suggest that during estrogen metabolism in the skin of the blackfin goodeid, ROS could be generated. During the hydroxylation of E_1 and E_2 carried out by CYP450 isoenzymes, estrogen-derived metabolites including 2-, 4-, and 16 α -hydroxyoestradiol (Bui and Weisz, 1989; Shou et al., 1997; Zhu and Conney, 1998; Chourasia and Joy, 2010) allow ROS generation (Cavaliere et al., 2000). The hydroxyl groups in a vicinal position on the 2- and 4-hydroxylated catechols could suffer further oxidation by semiquinones with consequent formation of O_2^* (Nutter et al., 1994; Tabakovic et al., 1996;

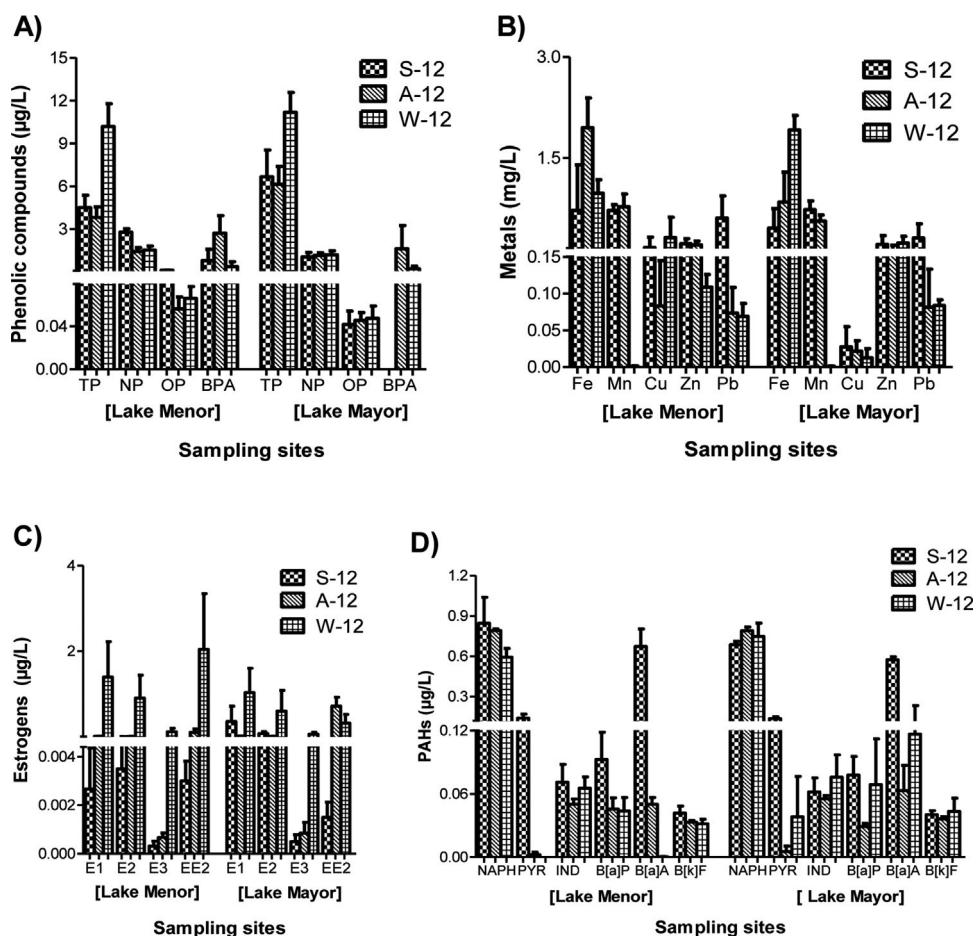


Fig. 6. Water levels of environmental pollutants in the lakes under study. Panel (A) phenolic compounds, TP, total phenols; NP, nonyl phenol; OP, octyl phenol; BPA, bisphenol A. Panel (B) metals. Panel (C) estrogens (E₁, estrone; E₂, 17- β -estradiol; E₃, estriol; EE₂, ethinyl estradiol). Panel (D) PAHs (IND, indeno[1,2,3-*c,d*]pyrene; NAPH, naphthalene; PYR, pyrene; B[a]A, benzo[a]anthracene; B[k]F, benzo[k]fluoranthene; B[a]P, benzo[a]pyrene).

Farinati et al., 2002; Roy et al., 2007; Okoh et al., 2011). The O₂[•] can achieve autodismutation by non-enzymatic via or dismutation by enzymatic via producing H₂O₂ (Hermes-Lima, 2004). Considering the role of CYP450 isoenzyme activities as a source of ROS and the alteration of CYP450 mRNAs by exposure to estrogenic compounds, we evaluated CYP 1A1 expression in the SML of wild blackfin goodeid, finding positive results. In the liver of *Carassius auratus* treated with EE₂, alterations in CYP 1A1 mRNA (Yan et al., 2012) and on CYP 1A1 protein expression in the liver of *Moxostoma hubbsi* (Maltais and Roy, 2014) were documented. A similar response was noted in male goldfish exposed to E₂ (Yan et al., 2013). CYP1A expression was also detected on the skin scales of *Carassius auratus* treated i.p. with β -naphthoflavone (Quirós et al., 2007) and in the scales of Atlantic salmon (*Salmo salar*) exposed to polychlorinated biphenyls (PCBs) and polyaromatic hydrocarbons (PAHs) (Rees et al., 2005; Van Beneden et al., 2010). The results of this study indicate that hydroxylation of exogenous estrogen in the SML mediated by CYP450 isoenzymes is likely to be a source of ROS. Differences with regard to male blackfin goodeids from a less estrogen-polluted lake (Lake Mayor) indicate that endogenous levels of these compounds in female fish from this lake can potentiate the toxicity of these endocrine disrupting compounds. However, these relationships were found only in the winter, suggesting that during the summer and autumn the concentration of estrogen in the water was below a threshold needed to induce CYP450 isoenzymes in the skin of the blackfin goodeid. In this regard, a concentration-dependent response was shown between E₂ concentrations in water and ROS generation in the liver of Japanese

sea bass *Lateolabrax japonicus* (Thilagam et al., 2010). These authors concluded that hepatic ROS levels might serve as a biomarker to indicate estrogen contamination. This response could be augmented by cross-talk between the estrogen receptor (ER) and the aryl hydrocarbon receptor (AhR) agonist (Andersson et al., 2007; Mortensen and Arukwe, 2007; Vega-López et al., 2008b; Gräns et al., 2010; Yan et al., 2012), as is the case with exogenous estrogens and PAHs. Findings of PCA and RDA showed that the metabolism of certain PAHs such as benzo[k]fluoranthene and indeno[1,2,3-*c,d*]pyrene was also a source of ROS in the skin of the female blackfin goodeid from Lake Mayor. Despite there being no preceding studies about ROS generation mediated by CYP450 isoenzymes in the SML of fish species, in fish liver a potential impairment of electron flow during the redox process of these isoenzymes may generate ROS in the presence of molecular oxygen (Schleizinger et al., 2006; Vega-López et al., 2009; Arzuaga and Elskus, 2010). Independent of the source of ROS in the SML of the wild blackfin goodeid of both sexes, the greater levels of ROS in wild fish than in control specimens confirms the usefulness of evaluating ROS in the SML of *G. viviparus* in monitoring the metabolism of exogenous estrogens and some PAHs carried out in the fish skin, in addition to other compounds such as water total phenols (TP).

Relations between ROS levels with TP suggest that these compounds are metabolized in the osteoblast and osteoclast scales of the blackfin goodeid, generating ROS, which were diffusible to the mucus layer. In this regard, phenol could produce hydroquinones mediated by the activity of liver CYP450 isoenzymes. Further, the hydroquinones could suffer redox reactions by NADPH-cytochrome

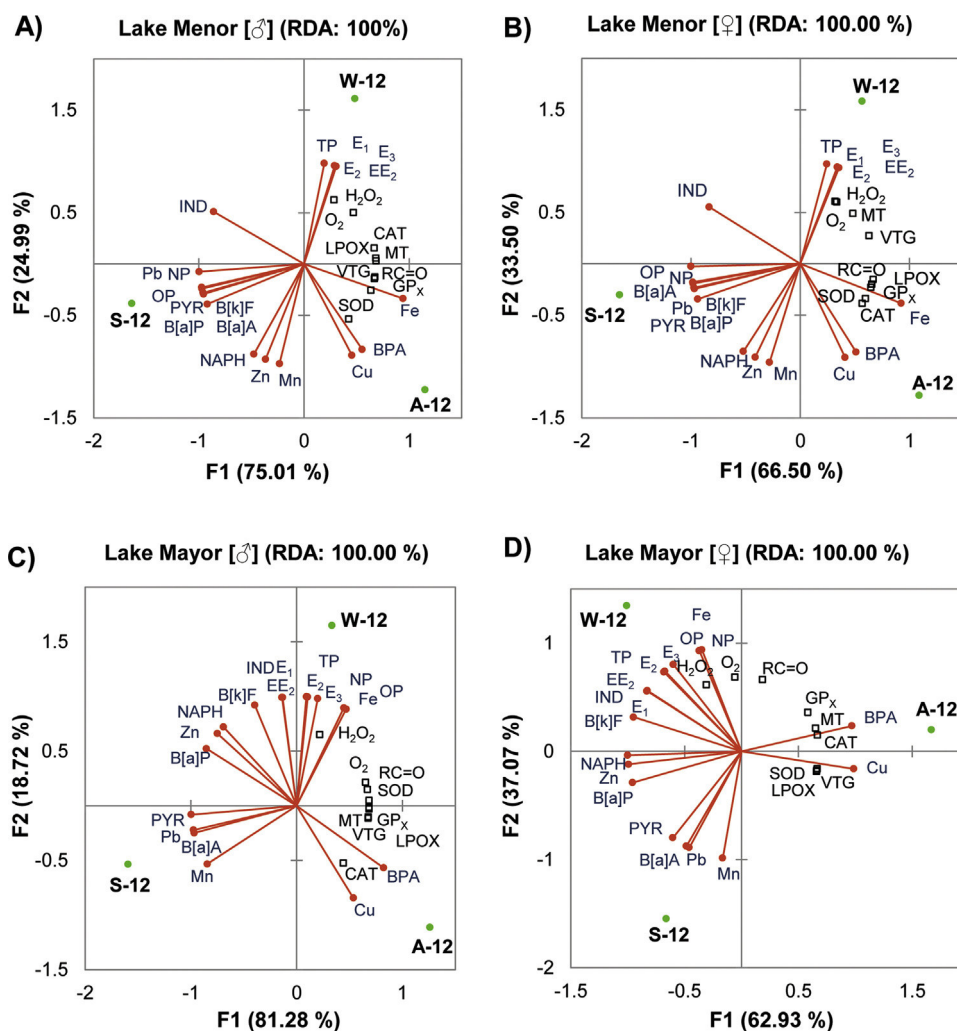


Fig. 7. Redundance analysis (RDA) showing the relationships between biological response variables: O_2^* , H_2O_2 , TBARS, $RC=O$, SOD, CAT, GPx, VTG and MT, and environmental data (explanatory variables: E_1 , estrone; E_2 , 17 β -estradiol; E_3 , estril; EE_2 , ethinyl estradiol; TP, total phenols; NP, nonyl phenol; OP, octyl phenol; BPA, bisphenol A; IND, indeno[1,2,3-*c,d*]pyrene; NAPH, naphthalene; PYR, pyrene; B[a]A, benzo[a]anthracene; B[k]F, benzo[k]fluoranthene; B[a]P, benzo[a]pyrene; Fe, Cu, Mn, Pb and Zn). In fish from Lake Menor the first bi-plot (RDA1) explained a high correlation between the response variables and explanatory variables, both in male (75.01%) and female fish (66.50%). In blackfin goodeid from Lake Mayor the ordination (RDA1) explained 81.28% and 66.50% of the variation in the data in male and female fish, respectively. The triplot (2-dimensional bi-plots; RDA1 and RDA2) showed 100% correlation between the response variables and explanatory variables. S-12: summer 2012; A-12: autumn 2012; W-12: winter 2012.

P450 reductase activity forming semiquinones, which under aerobic conditions may generate O_2^* (Joseph and Jaiswal, 1998; Roy et al., 2007; Okoh et al., 2011; Ramírez et al., 2013). Similar relationships were detected with nonyl phenol (NP) and octyl phenol (OP) only in the winter proposing that activities of enzymes involved in phenol metabolism in the SML of blackfin goodeid are subject to a self-regulation modulated by substrate concentration. By exposure to halogenated phenolic compounds ROS induction in other fish tissues was detected (Luo et al., 2008; Dong et al., 2009; Li et al., 2013). However, others studies under controlled conditions are needed to clarify these findings.

The oxidative stress response can be considered as a balance between pro-oxidant forces (i.e. O_2^* , H_2O_2), biomarkers of oxidative damage (i.e. lipid peroxidation and protein oxidation), the activities of enzymes involved in antioxidant defense (SOD, CAT, GPx), and the content of unspecific antioxidant biomolecules (i.e. Hermes-Lima, 2004; Valavanidis et al., 2006; Vega-López et al., 2007b). In this way, it is important to explore the interrelations among these factors in order to reach a better understanding of the biological response and its relationship with toxicants able to induce this damage.

The current study showed a sex-linked response of the antioxidant enzymes that was related with ROS levels in the SML of male blackfin goodeid. Despite the importance of gender related differences in these enzymes, only a few studies have documented this phenomenon (McFarland et al., 1999; Meyer et al., 2003; Vega-López et al., 2007b). In an earlier study assessing a complete panel of antioxidant defenses, CAT activity was higher in male than in female rats (Pinto and Bartley, 1969) as were seen in the goby (*Zosterisessor ophiocephalus*) (Livingstone et al., 1995). In male *G. viviparus* (Vega-López et al., 2007b) and in male *Ameioba splendens*, another member of the Goodeidae Family (Vega-López et al., 2009), hepatic SOD and CAT activities were greater than in female fish. These results are in agreement with the current study despite these studies being conducted in the liver; in contrast, in fresh plasma samples of the urodele amphibian *Pleurodeles waltli*, SOD and CAT activities were higher in females than in males (Alekhova et al., 2001). However, the relation to antioxidant defenses with ROS in the male fish SML could be indirect because some toxicants are able to induce oxidative stress on *G. viviparus* SML of both sexes.

During the seasons under study, the concentrations of toxic compounds able to oxidize lipids and proteins in the SML of

the blackfin goodeid showed fluctuations that were related with the biological response. In agreement with the current results, in *Acipenser ruthenus* spermatozoa an increase in SOD activity has been detected after exposure to BPA (Hulak et al., 2013), as in several mouse tissues (Kabuto et al., 2003). In contrast, diminution of these enzymes in zebrafish embryos was observed after short-term exposure to BPA (Wu et al., 2011), as was also the case in the testes of Sprague-Dawley rats (Chen et al., 2012). The results of the present study and preceding reports seem to be contradictory about the toxic effects of BPA elicited on antioxidant defenses; however, it is important to stress that the target tissue, species and exposure were not the same. Additionally, toxicants may overwhelm detoxification processes, particularly when the toxic effects involve an agent's adsorption, distribution, readsorption and toxication in the entire organism (Gregus, 2008). In this way, the relevance of the SML of fish in the context of an environmental monitoring program stands out because fish skin is a metabolically active site for certain compounds so it is unnecessary for all the processes cited above to occur.

It is well known that imbalances in electron flux during redox processing of metals (e.g. Fe and Cu) induce ROS by Fenton and Haber-Weiss reactions (Hermes-Lima, 2004). The relation of SOD and peroxidases in the SML of the blackfin goodeid with these divalent/polyvalent metals suggests that the high metabolism of the enzymes involved in antioxidant defenses is able to diminish ROS levels generated by redox processing of metals during the autumn compared to the winter. Similar results have been found in other tissues and fish species (Livingstone et al., 1995; McFarland et al., 1999; Meyer et al., 2003; Vega-López et al., 2012), even though the seasonality of the biological responses was not clearly documented.

In female fish SML the median Cu concentration in the water and the highest BPA content were linked with the highest TBARS levels and with RC=O in specimens of both sexes from Lake Menor as observed also in the SML of female fish from Lake Mayor during October. These results suggest a probable synergism between Cu and BPA or potentiating the effects of BPA. To our knowledge previous reports about Cu and BPA interactions on oxidative stress in fish do not exist; however, in some studies increased damage of this type was found in fish exposed to BPA. By the other hand, in the SML of Lake Mayor male specimens relationships of TBARS and RC=O with the NP and OP content of the water without seasonality were found. In sterlet (*Acipenser ruthenus*) spermatozoa (10^8 cells) treated with BPA or with NP an increase in TBARS and RC=O was observed from 2.5 to 10 µg/L (Hulak et al., 2013). Lipid peroxidation accompanied by decreases in SOD and CAT activities in a concentration-dependent pattern above 10 µg/L in zebrafish embryos exposed to BPA has also been found (Wu et al., 2011). The unfertilized eggs of Chinese rare minnow (*Gobiocypris rarus*) treated with NP from 50 to 200 µg/L showed a positive correlation between ROS, TBARS and protein carbonyl induction accompanied by SOD inhibition (Zhang et al., 2008). Oxidative damage elicited by NP and BPA in the blackfin goodeid can be explained by ROS generation and inhibition of antioxidant enzymes, as occurs in mammals (Choi et al., 2014). Using molecular docking it was documented that mammalian CAT and SOD were maximally inhibited by BPA and NP, respectively (Jayakanthan et al., 2015). However, during the summer the lower ROS content, TBARS and RC=O levels and low activities of antioxidant defenses in the SML of male blackfin goodeid from Lake Mayor coincided with the high NP concentration. These findings suggest that oxidative damage by exposure to NP and OP is a discrete process, probably affecting the fish when other alkyl phenols such as BPA reach threshold levels needed to induce oxidative stress. It stands out that the metabolism of phenolic compounds is surely a source of $O_2^{\bullet}$ and H_2O_2 in the SML of *G. viviparus* despite differences in the chemical structure of these

toxicants. In addition, the present results provide greater evidence about the environmental relevance of phenolic compounds and show a greater sensitivity of the skin of adult *G. viviparus* to oxidative stress elicited by alkyl phenols than did zebrafish embryos, sterlet spermatozoa and eggs of Chinese rare minnow. Further studies under controlled conditions are needed to reinforce or refute the current supposition about the role of phenol metabolism as a source of ROS and oxidative damage in the SML of *G. viviparus*.

Environmental levels of Fe and Cu contributed to lipid peroxidation and protein oxidation in the SML of the blackfin goodeid of both sexes. However, the greatest oxidative damage in the SML of Lake Menor specimens coincided with the higher levels of these metals found during the autumn, contrasting with a less differentiated seasonality in the SML of blackfin goodeid from Lake Mayor. The coincidence in the peaks of SML oxidative stress biomarkers from Lake Menor specimens with the highest heavy metal contents (Fe and Cu) suggest a true statistical relationship, contrasting with the findings of Lake Mayor specimens, where the observed relations could be considered a coincidence fact attributable to other toxic compounds as discussed above.

The role mixtures of metals play as well as environmental exposure in oxidative stress induction in the gill, liver and kidney of some fish species is well known (i.e. Oakes and Van Der Kraak, 2003; Valavanidis et al., 2006; Monserrat et al., 2007; Sevcikova et al., 2011; Mahboob, 2013). The current results evaluated as TBARS and RC=O in the SML of the blackfin goodeid are in agreement with previous results showing that this target is suitable for monitoring oxidative stress. However, there have been few reports about mucus production as a resistance mechanism in fish exposed to metals (McDonald and Wood, 1993; Andreji et al., 2005). The brown trout (*Salmo trutta*) treated with Cu produces a significant amount of mucus to reduce the uptake of Cu into the gills (Hansen et al., 2007). These authors proposed that acclimation to chronic metal exposure involve different strategies such as mucus production as well as the activation of metal-related stress gene transcription (metallothioneins). The concentration of sialic acid as a measurement of mucus production in the intestine of the rainbow trout (*Oncorhynchus mykiss*) pretreated with Zn in the diet was increased by a single dose of Cd (Khan and McGeer, 2013). However, in the liver of *Oreochromis* sp. exposed a significant induction of MT was detected; nevertheless, the density and size of producing-mucus cells on the gills were not changed (Wu et al., 2007). This result showed that there is not a direct correlation between MT and mucus content in fish exposed to metals. Regarding the possible role of MT in diminution of oxidative damage, the current results suggest that MT in the SML does not protect the skin of the blackfin goodeid against the oxidative stress elicited by metals because the peak of MT production in this fish species coincided with greater oxidative damage. However, the increase in mucus production in fish exposed to metals (Hansen et al., 2007; Khan and McGeer, 2013) is crucial considering the extent of the skin surface as the first target organ and by the role of the skin in innate immunity (Ellis, 2001; Tasumi et al., 2004; Tsutsui et al., 2005; De Veer et al., 2007; Esteban, 2012). This stresses the need for more studies to clarify the MT content in the mucus of fish exposed to metals.

For first time it was found by PCA and RDA that VTG levels in the SML of male blackfin goodeid from both lakes and female fish from Lake Menor were related to the concentration of BPA in the water during the autumn. To our knowledge there are no preceding reports about VTG induction in the SML of fish species by exposure to BPA; however, the estrogenicity of this compound is wide recognized (Larsen et al., 2006; Crain et al., 2007; Shanle and Xu, 2011). The estrogenicity of BPA has also been documented in the liver of females of this fish species using canonical correspondence analysis (Oliveras-Rubio et al., 2015). Vitellogenin, a phosphoglycolipoprotein precursor of oocyte yolk, is synthesized in the liver

of egg-laying vertebrates under the control of 17- β estradiol (Tata and Smith, 1979). Nevertheless, many sorts of toxicants can activate the estrogen receptor (ER) due to its high promiscuity (Shanle and Xu, 2011), inducing VTG synthesis (Van der Oost et al., 2003). VTG in the SML of male fish from Lake Mayor was related to the lower concentrations of NP and OP in the water as documented in the SML of other fish species (Meucci and Arukwe, 2005; Arukwe and Røe, 2008; Rey Vázquez et al., 2009; Maltais and Roy, 2014). The lack of a relationship of NP and OP concentrations in the water with VTG in the SML of male blackfin goodeid from Lake Menor and in female fish from both lakes probably confirms the discrete process of oxidation elicited by NP and OP affecting the fish response as discussed above.

Independent of the toxic relationships, after VTG synthesis and its post-transcriptional modification, this protein is released to the bloodstream to reach its target tissue, which in female fish is the oocyte (Tata and Smith, 1979). During the circulation of VTG in the bloodstream it is possible that this protein can reach the SML due to the elevated vasculature in the skin; however, previous studies about VTG transfer to fish skin do not exist. Despite the foregoing it has been proposed that VTG confinement in mucus vacuoles of the skin serves as an excretory pathway in male fish (Moncaut et al., 2003). Nevertheless, in the SML of female fish VTG could have a pheromonal function as has been proposed for female garter snakes (Garstka and Crews, 1981), as in the South American cichlid fish *Cichlasoma dimerus* (Moncaut et al., 2003).

Interestingly, VTG levels in the SML were related with the Fe and Cu content of the water, both in male and female blackfin goodeid, using PCA or RDA. However, the relationships apparently indicate that these metals potentiate the endocrine disrupting effects of alkylphenols and estrogens. Nevertheless, further studies under controlled conditions are required on this topic.

The results of the current study showed a larger condition factor (K) for the Lake Menor specimens with regard to fish from Lake Mayor. This response probably could be explained by the amount of available food in Lake Menor, where a greater diversity of phytoplankton was reported (Vega-López et al., 2013). It is considered that a K value greater than 1.0 denotes a better condition of the fish (Le Cren, 1951; Jin et al., 2015). Although this is true for practical purposes in fisheries (Jennings et al., 2014), anthropogenic alterations could decrease the body size of fishes (Audzijonyte et al., 2013). In a program to monitor fish health as in our study, the K value does not reflect the impact of toxicants on the fish skin of both sexes of blackfin goodeid. Another feature commonly used to describe a fish population is its weight (Jin et al., 2015). Male fish from Lake Mayor attained a greater weight, which was accompanied by low levels of damage in the SML regarding male fish from Lake Menor. A comparison between the weights of fish with the biomarkers evaluated in the SML did not provide confident findings because this factor could be influenced by the liver weight.

5. Conclusion

The current results corroborate that the skin mucus layer (SML) of wild *G. viviparus* is a sensitive target for monitoring the impacts of complex mixtures of toxicants found in aquatic ecosystems from the Valley of Mexico. The metabolism of estrogenic and phenolic compounds such as alkylphenols is involved with the biological response in the skin of this fish species, possibly mediated by ROS generation. This statement is based on the number of the relationships between ROS ($O_2^{\bullet}$, H_2O_2), oxidative stress biomarkers and the activities of enzymes involved in antioxidant defense such as SOD, CAT and GPx. In this regard, it is possible to propose the following toxicological order of importance: BPA > E_1 , E_2 , E_3 , EE2 > total phenols > NP and OP in addition to Fe and Cu. VTG content in the SML

of the blackfin goodeid showed a sex-linked response, i.e. in male fish levels of this biomolecule were influenced by BPA during the autumn and apparently potentiated by Cu and Fe. In contrast, in the SML of female fish, the endocrine disrupting effect was due to the influence of estrogens during the winter. On the other hand, MT was correlated with Fe and Cu; however, it was not possible to affirm that MT in the SML played a protective role in reducing oxidative stress elicited by metals. What stands out is a sex-linked response in the SML of *G. viviparus* that depends on the nature of the toxicants, their concentrations, bioavailability and interactions, in addition to the season of the year. Further studies under controlled conditions are needed to clarify these assumptions with the aim of validating this approach to monitoring of fish health.

Acknowledgements

This study was supported by Instituto Politécnico Nacional-SIP code 20130795 and CONACyT-FOMIX. R. Dzúl-Caamal, H. F. Olivares-Rubio, and M. A. Rocha-Gómez are DSc. Students which receives scholarship from CONACyT and BEIFI-IPN. L. Salazar-Coria is a DSc. student. A. Vega-López is fellow of Estímulos al Desempeño en Investigación and Comisión y Fomento de Actividades Académicas (Instituto Politécnico Nacional) and Sistema Nacional de Investigadores (SNI, CONACyT, México).

References

- Alekshova, T., Sof'in, A., Kobelkova, T., Marco, R., Dournon, C., 2001. Sex-linked differences in activity of enzymes in the blood of the urodele amphibian *Pleurodeles waltl*. *Comp. Biochem. Physiol. A* 130, 819–825.
- Andersson, C., Katsiadaki, I., Lundstedt-Enkel, K., Orberg, J., 2007. Effects of 17 α -ethynylestradiol on EROD activity, spiggin and vitellogenin in three-spined stickleback (*Gasterosteus aculeatus*). *Aquat. Toxicol.* 83, 33–42.
- Andreji, J., Stranai, I., Masanyi, P., Valent, M., 2005. Concentration of selected metals in muscle of various fish species. *J. Environ. Sci. Health A: Tox. Hazard. Subst. Environ. Eng.* 40, 899–912.
- APHA, 1998. Standard Methods for Examination of Water and Waste Water, 20th ed. American Public Health Association, Washington, DC.
- Arukwe, A., Røe, K., 2008. Molecular and cellular detection of expression of vitellogenin and zona radiata protein in liver and skin of juvenile salmon (*Salmo salar*) exposed to nonylphenol. *Cell Tissue Res.* 331, 701–712.
- Arzuaga, X., Elskus, A., 2010. Polluted-site killifish (*Fundulus heteroclitus*) embryos are resistant to organic pollutant-mediated induction of CYP1A activity, reactive oxygen species, and heart deformities. *Environ. Toxicol. Chem.* 29, 676–682.
- Audzijonyte, A., Kuparinen, A., Gorton, R., Fulton, E.A., 2013. Ecological consequences of body size decline in harvested fish species: positive feedback loops in trophic interactions amplify human impact. *Biol. Lett.* 9, 20121103.
- Benhamed, S., Guardiola, F.A., Mars, M., Esteban, M.A., 2014. Pathogen bacteria adhesion to skin mucus of fishes. *Vet. Microbiol.* 171, 1–12.
- Ben-Khedher, S., Jebali, J., Kamel, N., Banni, M., Rameh, M., Irad, A., Boussetta, H., 2013. Biochemical effects in crabs (*Carcinus maenas*) and contamination levels in the Bizerta Lagoon: an integrated approach in biomonitoring of marine complex pollution. *Environ. Sci. Pollut. Res.* 20, 2616–2631.
- Buege, J.A., Aust, S.D., 1978. Microsomal lipid peroxidation. *Methods Enzymol.* 52, 302–310.
- Bui, Q.D., Weisz, J., 1989. Monooxygenase mediating catecholesterogen formation by rat anterior pituitary is an estrogen-4-hydroxylase. *Endocrinology* 124, 1085–1087.
- Cavalieri, E., Frenkel, K., Liehr, J.G., Rogan, E., Roy, D., 2000. Estrogens as endogenous genotoxic agents – DNA adducts and mutations. *J. Natl. Cancer Inst. Monogr.* 27, 75–93.
- Chen, M., Xu, B., Ji, W., Qiao, S., Hu, N., Hu, Y., Wu, W., Qiu, L., Zhang, R., Wang, Y., Wang, S., Zhou, Z., Xia, Y., Wang, X., 2012. Bisphenol A alters n-6 fatty acid composition and decreases antioxidant enzyme levels in rat testes: a LC-QTOF-based metabolomics study. *PLoS ONE* 7, e44754.
- Choi, M.S., Park, H.J., Oh, J.H., Lee, E.H., Park, S.M., Yoon, S., 2014. Nonylphenol-induced apoptotic cell death in mouse TM4 Sertoli cells via the generation of reactive oxygen species and activation of the ERK signaling pathway. *J. Appl. Toxicol.* 34, 628–636.
- Chourasia, T.K., Joy, K.P., 2010. Seasonal variation in tissue estrogen-2/4-hydroxylases (EH) and in vitro effects of steroids on ovarian EH activity in the catfish *Heteropneustes fossilis*. *Steroids* 75, 1097–1105.
- Cone, R.A., 2009. Barrier properties of mucus. *Adv. Drug Deliv. Rev.* 61, 75–85.
- Crain, D.A., Eriksen, M., Iguchi, T., Jobling, S., Laufer, H., LeBlanc, G.A., Guillette, L.J., 2007. An ecological assessment of bisphenol-A: evidence from comparative biology. *Reprod. Toxicol.* 24, 225–239.
- De Veer, M.J., Kemp, J.M., Meeusen, E.N., 2007. The innate host defence against nematode parasites. *Parasite Immunol.* 29, 1–9.

- Dong, Y.L., Zhou, P.J., Jiang, S.Y., Pan, X.W., Zhao, X.H., 2009. Induction of oxidative stress and apoptosis by pentachlorophenol in primary cultures of *Carassius carassius* hepatocytes. *Comp. Biochem. Physiol. C* 150, 179–185.
- Dzul-Caamal, R., Domínguez-López, M.L., García-Latorre, E., Vega-López, A., 2012. Implications of cytochrome 450 isoenzymes, aryl-esterase and oxonase activity in the inhibition of the acetylcholinesterase of *Chiostoma jordani* treated with phosphorothionate pesticides. *Ecotoxicol. Environ. Saf.* 84, 199–206.
- Dzul-Caamal, R., Olivares-Rubio, H.F., López-Tapia, P., Vega-López, A., 2013. Pro-oxidant and antioxidant response elicited by CH_2Cl_2 , CHCl_3 and BrCHCl_2 in *Goodea gracilis* using non-invasive methods. *Comp. Biochem. Physiol. A* 165, 515–527.
- Ellis, A.E., 2001. Innate host defense mechanisms of fish against viruses and bacteria. *Dev. Comp. Immunol.* 25, 827–839.
- Esteban, M., Castaño, A., 2009. Non-invasive matrices in human biomonitoring: a review. *Environ. Int.* 35, 438–449.
- Esteban, M.A., 2012. An overview of the immunological defenses in fish skin. *ISRN Immunol.* 2012, 29.
- Farinati, F., Cardin, R., Bortolami, M., Grottola, A., Manno, M., Colantoni, A., Villa, E., 2002. Estrogens receptors and oxidative damage in the liver. *Mol. Cell Endocrinol.* 193, 85–88.
- Garshelis, D.L., 2006. On the allure of noninvasive genetic sampling—putting a face to the name. *Ursus* 17, 109–123.
- Garstka, W.R., Crews, D., 1981. Female sex pheromone in the skin and circulation of a garter snake. *Science* 214, 681–682.
- Gregus, Z., 2008. Mechanisms of toxicity. In: Klaassen, C.D. (Ed.), Casarett & Doull's Toxicology: The Basic Science of Poisons. McGraw-Hill, New York, pp. 45–106.
- Gräns, J., Wassmur, B., Celander, M.C., 2010. One-way inhibiting cross-talk between arylhydrocarbon receptor (AhR) and estrogen receptor (ER) signaling in primary cultures of rainbow trout hepatocytes. *Aquat. Toxicol.* 100, 263–270.
- Hansen, B.H., Garmo, O.A., Olsvik, P.A., Andersen, R.A., 2007. Gill metal binding and stress gene transcription in brown trout (*Salmo trutta*) exposed to metal environments: the effect of pre-exposure in natural populations. *Environ. Toxicol. Chem.* 26, 944–953.
- Hermes-Lima, M., 2004. Oxygen in biology and biochemistry: role of free radicals. In: Storey, K.B. (Ed.), Functional Metabolism: Regulation and Adaptation. Wiley-Liss, Hoboken, pp. 319–368.
- Hulak, M., Gazo, I., Shalutin, A., Linhartova, P., 2013. In vitro effects of bisphenol A on the quality parameters, oxidative stress, DNA integrity and adenosine triphosphate content in sterlet (*Acipenser ruthenus*) spermatozoa. *Comp. Biochem. Physiol. C* 158, 64–71.
- Jayakanthan, M., Jubendradass, R., D'Cruz, S.C., Mathur, P.P., 2015. A use of homology modeling and molecular docking methods: to explore binding mechanisms of nonylphenol and bisphenol A with antioxidant enzymes. *Methods Mol. Biol.* 1268, 273–289.
- Jennings, S., Smith, A.D., Fulton, E.A., Smith, D.C., 2014. The ecosystem approach to fisheries: management at the dynamic interface between biodiversity conservation and sustainable use. *Ann. N. Y. Acad. Sci.* 1322, 48–60.
- Jin, S., Yan, X., Zhang, H., Fan, W., 2015. Weight-length relationships and Fulton's condition factors of skipjack tuna (*Katsuwonus pelamis*) in the western and central Pacific Ocean. *PeerJ* 3, e758.
- Joseph, P., Jaiswal, A.K., 1998. NAD(P)H:quinone oxidoreductase 1 reduces the mutagenicity of DNA caused by NADPH:P450 reductase-activated metabolites of benzo(a)pyrene quinones. *Br. J. Cancer* 77, 709–719.
- Kabuto, H., Hasuike, S., Minagawa, N., Shishibori, T., 2003. Effects of bisphenol A on the metabolisms of active oxygen species in mouse tissues. *Environ. Res.* 93, 31–35.
- Khan, F.R., McGeer, J.C., 2013. Zn-stimulated mucus secretion in the rainbow trout (*Oncorhynchus mykiss*) intestine inhibits Cd accumulation and Cd-induced lipid peroxidation. *Aquat. Toxicol.* 142–143, 17–25.
- Larsen, B.K., Bjørnstad, A., Sundt, R.C., Taban, I.C., Pampanin, D.M., Andersen, O.K., 2006. Comparison of protein expression in plasma from nonylphenol and bisphenol-A exposed Atlantic cod (*Gadus morhua*) and turbot (*Scophthalmus maximus*) by use of SELDI-TOF. *Aquat. Toxicol.* 78, S25–S33.
- Le Cren, E.D., 1951. The length-weight relationship and seasonal cycle in gonad weight and condition in Perch (*Perca fluviatilis*). *J. Anim. Ecol.* 20, 201–219.
- Lehane, D.B., McKie, N., Russell, R.G., Henderson, I.W., 1999. Cloning of a fragment of the osteonectin gene from goldfish, *Carassius auratus*: its expression and potential regulation by estrogen. *Gen. Comp. Endocrinol.* 114, 80–87.
- Lei, X.G., Evenson, J.K., Thompson, K.M., Sunde, R.A., 1995. Glutathione peroxidase and phospholipid hydroperoxide glutathione peroxidase are differentially regulated in rats by dietary selenium. *J. Nutr.* 125, 1438–1446.
- Levine, R.L., Williams, J.A., Stadtman, E.P., Shacter, E., 1994. Carbonyl assays for determination of oxidatively modified proteins. *Methods Enzymol.* 233, 346–357.
- Li, H., Zhang, X., Qiu, Q., An, Z., Qi, Y., Huang, D., Zhang, Y., 2013. 2,4-Dichlorophenol induces apoptosis in primary hepatocytes of grass carp (*Ctenopharyngodon idella*) through mitochondrial pathway. *Aquat. Toxicol.* 140–141, 117–122.
- Livingstone, D.R., Lemaire, P., Matthews, A., Peters, L.D., Porte, C., Fitzpatrick, P.J., Forlin, L., Nasci, C., Fossatto, V., Wootton, N., Goldfarb, P., 1995. Assessment of the impact of organic pollutants on Goby (*Zosterisessor ophicephalus*) and mussel (*Mytilus galloprovincialis*) from the Venice Lagoon, Italy: biochemical studies. *Mar. Environ. Res.* 39, 235–240.
- Luo, Y., Sui, Y.X., Wang, X.R., Tian, Y., 2008. 2-Chlorophenol induced hydroxyl radical production in mitochondria in *Carassius auratus* and oxidative stress – an electron paramagnetic resonance study. *Chemosphere* 71, 1260–1268.
- Mackay, P., Zielinski, W.J., Long, R.A., Ray, J.C., 2008. Noninvasive research and carnivore conservation. In: Long, R.A., Mackay, P., Zielinski, W.L., Ray, J.C. (Eds.), Noninvasive Survey Methods for Carnivores. Island Press, Washington, DC, pp. 1–7.
- Macpherson, A.J., Geuking, M.B., McCoy, K.D., 2005. Immune responses that adapt the intestinal mucosa to commensal intestinal bacteria. *Immunology* 115, 153–162.
- Mahboob, S., 2013. Environmental pollution of heavy metals as a cause of oxidative stress in fish: a review. *Life Sci.* 10, 10(10s).
- Maltais, D., Roy, R.L., Couillard, C.M., 2010. Hybrid ELISAs for vitellogenins of the endangered copper redhorse *Moxostoma hubbsi* and the shorthead redhorse *Moxostoma macrolepidotum* (Cypriniformes, Catostomidae). *Ecotoxicol. Environ. Saf.* 73, 883–892.
- Maltais, D., Roy, R.L., 2014. Effects of nonylphenol and ethinylestradiol on copper redhorse (*Moxostoma hubbsi*), an endangered species. *Ecotoxicol. Environ. Saf.* 108, 168–178.
- McDonald, D.C., Wood, C.M., 1993. Branchial mechanism of acclimation to metals in freshwater fish. In: Rankin, C., Jensen, F.B. (Eds.), Fish Ecophysiology. Chapman & Hall, London, pp. 295–319.
- McFarland, V.A., Inouye, L.S., Lutz, C.H., Jarvis, A.S., Clarke, J.U., McCant, D.D., 1999. Biomarkers of oxidative stress and genotoxicity in livers of field-collected brown bullhead, *Ameiurus nebulosus*. *Arch. Environ. Contam. Toxicol.* 37, 236–241.
- Meucci, V., Arukwe, A., 2005. Detection of vitellogenin and zona radiata protein expressions in surface mucus of immature juvenile Atlantic salmon (*Salmo salar*) exposed to waterborne nonylphenol. *Aquat. Toxicol.* 73, 1–10.
- Meyer, J.N., Smith, J.D., Winston, G.W., Di Giulio, R.T., 2003. Antioxidant defenses in killifish (*Fundulus heteroclitus*) exposed to contaminated sediments and model prooxidants: short-term and heritable responses. *Aquat. Toxicol.* 65, 377–395.
- Misra, H.P., Fridovich, I., 1972. The role of superoxide dismutase anion in the autooxidation of epinephrine and a simple assay for superoxide dismutase. *J. Biol. Chem.* 247, 3170–3175.
- Moncaut, N., Nostro, F.L., Maggese, M.C., 2003. Vitellogenin detection in surface mucus of the South American cichlid fish *Cichlasoma dimerus* (Heckel, 1840) induced by estradiol-17beta. Effects on liver and gonads. *Aquat. Toxicol.* 63, 127–137.
- Monserat, J.M., Martínez, P.E., Geracitano, L.A., Amado, L.L., Martins, C.M., Pinho, G.L., Chaves, I.S., Ferreira-Cravo, M., Ventura-Lima, J., Bianchini, A., 2007. Pollution biomarkers in estuarine animals: critical review and new perspectives. *Comp. Biochem. Physiol. C* 146, 221–234.
- Mortensen, A.S., Arukwe, A., 2007. Effects of 17alpha-ethynylestradiol on hormonal responses and xenobiotic biotransformation system of Atlantic salmon (*Salmo salar*). *Aquat. Toxicol.* 85, 113–123.
- Needham, L.L., Ozkaynak, H., Whyatt, R.M., Barr, D.B., Wang, R.Y., Naeher, L., Akland, G., Bahadori, T., Bradman, A., Fortmann, R., Liu, L.J., Morandi, M., O'Rourke, M.K., Thomas, K., Quackenboss, J., Ryan, P.B., Zartarian, V., 2005. Exposure assessment in the National Children's study: introduction. *Environ. Health Perspect.* 113, 1076–1082.
- Nigam, A.K., Srivastava, N., Rai, A.K., Kumari, U., Mittal, A.K., Mittal, S., 2012. The first evidence of cholinesterases in skin mucus of carps and its applicability as biomarker of organophosphate exposure. *Environ. Toxicol.* 29, 788–796.
- Nigam, A.K., Kumari, U., Mittal, S., Mittal, A.K., 2014. Characterization of carboxylesterase in skin mucus of *Cirrhinus mrigala* and its assessment as biomarker of organophosphate exposure. *Fish Physiol. Biochem.* 40, 635–644.
- Nogueira, P., Pacheco, M., Pereira, M.L., Mendo, S., Rotchell, J.M., 2010. Novel potential molecular biomarkers of aquatic contamination in *Dicentrarchus labrax* and *Liza aurata*. In: Hamamura, N., Suzuki, S., Mendo, S., Barroso, C.M., Iwata, H., Tanabe, S. (Eds.), Interdisciplinary Studies on Environmental Chemistry—Biological Responses to Contaminants. Terrapub, pp. 127–138.
- NOM-059-SEMARNAT-2010, 2010. Norma Oficial Mexicana, Protección ambiental-especies nativas de México de flora y fauna silvestres categorías de riesgo y especificaciones para su inclusión, exclusión o cambio-lista de especies en riesgo, pp. 78.
- Nussey, G., Van Vuren, J.H.J., Du Preez, H.H., 2000. Bioaccumulation of chromium, manganese, nickel and lead in the tissues of the moggell, *Labeo umbratus* (Cyprinidae), from Witbank Dam, Mpumalanga. *Water SA* 26, 269–284.
- Nutter, L.M., Wu, Y.Y., Ngo, E.O., Sierra, E.E., Gutierrez, P.L., Abul-Hajj, Y.J., 1994. An o-quinone form of estrogen produces free radicals in human breast cancer cells: correlation with DNA damage. *Chem. Res. Toxicol.* 7, 23–28.
- Oakes, K.D., Van Der Kraak, G.J., 2003. Utility of the TBARS assay in detecting oxidative stress in white sucker (*Catostomus commersoni*) populations exposed to pulp mill effluent. *Aquat. Toxicol.* 63, 447–463.
- Okoh, V., Deoraj, A., Roy, D., 2011. Estrogen-induced reactive oxygen species-mediated signalings contribute to breast cancer. *Biochim. Biophys. Acta* 1815, 115–133.
- Olivares-Rubio, H.F., Dzul-Caamal, R., Gallegos-Rangel, M.E., Madera-Sandoval, R.L., Domínguez-López, M.L., García-Latorre, E., Vega-López, A., 2015. Relationship between biomarkers and endocrine-disrupting compounds in wild *Girardinichthys viviparus* from two lakes with different degrees of pollution. *Ecotoxicology* 24, 664–685.
- Pedersen, K.L., Pedersen, S.N., Højrup, P., Andersen, J.S., Roepstorff, P., Knudsen, J., Depledge, M.H., 1994. Purification and characterization of a cadmium-induced metallothionein from the shore crab *Carcinus maenas* (L.). *Biochem. J.* 297, 609–614.
- Pinto, R.E., Bartley, W., 1969. The nature of the sex-linked differences in glutathione peroxidase activity and aerobic oxidation of glutathione in male and female rat liver. *Biochem. J.* 115, 449–456.
- Quirós, L., Raldúa, D., Navarro, A., Casado, M., Barceló, D., Piña, B., 2007. A noninvasive test of exposure to toxicants: quantitative analysis of cytochrome P4501A expression in fish scales. *Environ. Toxicol. Chem.* 26, 2179–2186.

- Radi, R., Turrens, J.F., Chang, L.Y., Bush, K.M., Crapo, J.D., Freeman, B.A., 1991. [Detection of catalase in rat heart mitochondria](#). *J. Biol. Chem.* 266, 22028–22034.
- Ramírez, C., Pham, K., Franco, M.F., Chwa, M., Limb, A., Kuppermann, B.D., Kenney, M.C., 2013. [Hydroquinone induces oxidative and mitochondrial damage to human retinal Müller cells \(MIO-M1\)](#). *Neurotoxicology* 39, 102–108.
- Rees, C.B., McCormick, S.D., Li, W., 2005. [A non-lethal method to estimate CYP1A expression in laboratory and wild Atlantic salmon \(*Salmo salar*\)](#). *Comp. Biochem. Physiol. C* 141, 217–224.
- Rey Vázquez, G., Meijide, F., Da Cuna, R., Lo Nostro, F., Piazza, Y., Babay, P., Trudeau, V., Maggese, C., Guerrero, G., 2009. [Exposure to waterborne 4-tert-octylphenol induces vitellogenin synthesis and disrupts testis morphology in the South American freshwater fish *Cichlasoma dimerus* \(Teleostei, Perciformes\)](#). *Comp. Biochem. Physiol. C* 150, 298–306.
- Roy, D., Cai, Q., Felt, Q., Narayan, S., 2007. [Estrogen-induced generation of reactive oxygen and nitrogen species, gene damage, and estrogen-dependent cancers](#). *J. Toxicol. Environ. Health B: Crit. Rev.* 10, 235–257.
- SAGARPA, 2001. Norma Oficial Mexicana NOM-062-ZOO-1999. Especificaciones técnicas para la producción, cuidado y el uso de animales de laboratorio. Diario Oficial de la Federación, Segunda Sección, México, pp. 1–57.
- Schleizinger, J.J., Struntz, W.D., Goldstone, J.V., Stegeman, J.J., 2006. [Uncoupling of cytochrome P450 1A and stimulation of reactive oxygen species production by co-planar polychlorinated biphenyl congeners](#). *Aquat. Toxicol.* 77, 422–432.
- Sevcikova, M., Modra, H., Slaninova, A., Svobodova, Z., 2011. [Metals as a cause of oxidative stress in fish: a review](#). *Vet. Med. (Czech)* 56, 537–546.
- Shanle, E.K., Xu, W., 2011. [Endocrine disrupting chemicals targeting estrogen receptor signaling: identification and mechanisms of action](#). *Chem. Res. Toxicol.* 24, 6–19.
- Shou, M., Korzekwa, K.R., Brooks, E.N., Kraus, K.W., Gonzalez, F.J., Gelboin, H.V., 1997. [Role of human hepatic cytochrome P450 1A2 and 3A4 in the metabolic activation of estrone](#). *Carcinogenesis* 18, 207–214.
- Tabakovic, K., Gleason, W.B., Ojala, W.H., Abul-Hajj, Y.J., 1996. [Oxidative transformation of 2 hydroxyestrone. Stability and reactivity of 2,3-estrone quinone and its relationship to estrogen carcinogenicity](#). *Chem. Res. Toxicol.* 9, 860–865.
- Tasumi, S., Yang, W.J., Usami, T., Tsutsui, S., Ohira, T., Kawazoe, I., Wilder, M.N., Aida, K., Suzuki, Y., 2004. [Characteristics and primary structure of a galectin in the skin mucus of the Japanese eel, *Anguilla japonica*](#). *Dev. Comp. Immunol.* 28, 325–335.
- Tata, J.R., Smith, D.F., 1979. [Vitellogenesis: a versatile model for hormonal regulation of gene expression](#). *Rec. Prog. Horm. Res.* 35, 47–93.
- Ter Braak, C.J.F., Prentice, I.C., 1988. [A theory of gradient analysis](#). *Adv. Ecol. Res.* 18, 271–313.
- Thilagam, H., Gopalakrishnan, S., Qu, H.D., Bo, J., Wang, K.J., 2010. [17β estradiol induced ROS generation, DNA damage and enzymatic responses in the hepatic tissue of Japanese sea bass](#). *Ecotoxicology* 19, 1258–1267.
- Tlili, S., Jebali, J., Banni, M., Haouas, Z., Mlayah, A., Helal, A.N., Boussetta, H., 2010. [Multimarker approach analysis in common carp *Cyprinus carpio* sampled from three freshwater sites](#). *Environ. Monit. Assess.* 168, 285–298.
- Trenzado, C., Hidalgo, M.C., García-Gallego, M., Morales, A.E., Furné, M., Domezain, A., Domezain, J., Sanz, A., 2006. [Antioxidant enzymes and lipid peroxidation in sturgeon *Acipenser naccarii* and trout *Oncorhynchus mykiss*. A comparative study](#). *Aquaculture* 254, 758–767.
- Tsutsui, S., Tasumi, S., Suetake, H., Kikuchi, K., Suzuki, Y., 2005. [Demonstration of the mucosal lectins in the epithelial cells of internal and external body surface tissues in pufferfish \(*Fugu rubripes*\)](#). *Dev. Comp. Immunol.* 29, 243–253.
- Valavanidis, A., Vlahogianni, T., Dassenakis, M., Scoullas, M., 2006. [Molecular biomarkers of oxidative stress in aquatic organisms in relation to toxic environmental pollutants](#). *Ecotoxicol. Environ. Saf.* 64, 178–189.
- Van Beneden, R., Ashworth, S.L., Elskus, A., Gosse, J.A., Perkins, B., 2010. [Fish Scales as Non-Lethal Biosensors of Surface Water Contaminants](#). Report as of FY2010 for 2010ME214B, pp. 17.
- Van den Brink, P.J., van den Brink, N.W., Ter Braak, C.J.F., 2003. [Multivariate analysis of ecotoxicological data using ordination: demonstration of utility on the basis of various examples](#). *Aust. J. Ecotoxicol.* 9, 141–156.
- Van der Oost, R., Beyer, J., Vermeulen, N.P.E., 2003. [Fish bioaccumulation and biomarkers in environmental risk assessment: a review](#). *Environ. Toxicol. Pharmacol.* 13, 57–149.
- Vega-López, A., Martínez-Tabche, L., Domínguez-López, M.L., García-Latorre, E., Ramón-Gallegos, E., García-Gasca, A., 2006. [Vitellogenin induction in the endangered goodeid fish *Girardinichthys viviparus*: vitellogenin characterization and estrogenic effects of polychlorinated biphenyls](#). *Comp. Biochem. Physiol. C* 142, 356–364.
- Vega-López, A., Ortiz-Ordóñez, E., Uría-Galicia, E., Mendoza-Santana, E.L., Hernández-Cornejo, R., Atondo-Mexia, R., García-Gasca, A., García-Latorre, E., Domínguez-López, M.L., 2007a. [The role of vitellogenin during gestation of *Girardinichthys viviparus* and *Ameba splendens*: two goodeid fish with matrotrophic viviparity](#). *Comp. Biochem. Physiol. A* 147, 731–742.
- Vega-López, A., Galar-Martínez, M., Jiménez-Orozco, F.A., García-Latorre, E., Domínguez-López, M.L., 2007b. [Gender related differences in the oxidative stress response to PCB exposure in an endangered goodeid fish \(*Girardinichthys viviparus*\)](#). *Comp. Biochem. Physiol. A* 146, 672–678.
- Vega-López, A., Jiménez-Orozco, F.A., García-Latorre, E., Domínguez-López, M.L., 2008a. [Oxidative stress response elicited in an endangered goodeid fish \(*Girardinichthys viviparus*\) by water from its type localities](#). *Ecotoxicol. Environ. Saf.* 71, 94–103.
- Vega-López, A., Jiménez-Orozco, F.A., Ramón-Gallegos, E., García-Latorre, E., Domínguez-López, M.L., 2008b. [Estrogenic effects of polychlorinated biphenyls and relation to cytochrome P4501A activity in the endangered goodeid fish *Ameba splendens*](#). *Environ. Toxicol. Chem.* 27, 963–969.
- Vega-López, A., Jiménez-Orozco, F.A., Jiménez-Zamudio, L.A., García-Latorre, E., Domínguez-López, M.L., 2009. [Prooxidant and antioxidant sex-linked response and its relationship to mixed oxidase function enzymes in liver of *Ameba splendens*, an endangered goodeid fish exposed to PCBs](#). *Toxicol. Environ. Chem.* 91, 315–330.
- Vega-López, A., Carrillo-Morales, C.I., Olivares-Rubio, H.F., Domínguez-López, M.L., García-Latorre, E.A., 2012. [Evidence of bioactivation of halomethanes and its relation to oxidative stress response in *Chirostoma riojai*, an endangered fish from a polluted lake in Mexico](#). *Arch. Environ. Contam. Toxicol.* 62, 479–493.
- Vega-López, A., Ayala-López, G., Posadas-Espadas, B.P., Olivares-Rubio, H.F., Dzul-Caamal, R., 2013. [Relations of oxidative stress in freshwater phytoplankton with heavy metals and polycyclic aromatic hydrocarbons](#). *Comp. Biochem. Physiol. A* 165, 498–507.
- Wang, L., Cai, Y.Q., He, B., Yuan, C.G., Shen, D.Z., Shao, J., Jiang, G.B., 2006. [Determination of estrogens in water by HPLC-UV using cloud point extraction](#). *Talanta* 1, 47–51.
- Wu, S.M., Shih, M.J., Ho, Y.C., 2007. [Toxicological stress response and cadmium distribution in hybrid tilapia \(*Oreochromis sp.*\) upon cadmium exposure](#). *Comp. Biochem. Physiol. C* 145, 218–226.
- Wu, M., Xu, H., Shen, Y., Qiu, W., Yang, M., 2011. [Oxidative stress in zebrafish embryos induced by short-term exposure to bisphenol A, nonylphenol, and their mixture](#). *Environ. Toxicol. Chem.* 30, 2335–2341.
- Yan, Z., Lu, G., He, J., 2012. [Reciprocal inhibiting interactive mechanism between the estrogen receptor and aryl hydrocarbon receptor signaling pathways in goldfish \(*Carassius auratus*\) exposed to 17β-estradiol and benzo\[a\]pyrene](#). *Comp. Biochem. Physiol. C* 156, 17–23.
- Yan, Z., Lu, G., Wu, D., Ye, Q., Xie, Z., 2013. [Interaction of 17β-estradiol and ketoconazole on endocrine function in goldfish \(*Carassius auratus*\)](#). *Aquat. Toxicol.* 132–133, 19–25.
- Zhang, X., Yang, F., Cai, Y.Q., Xu, Y., 2008. [Oxidative damage in unfertilized eggs of Chinese rare minnow \(*Gobiocypris rarus*\) exposed to nonylphenol](#). *Environ. Toxicol. Chem.* 27, 213–219.
- Zhu, B.T., Conney, A.H., 1998. [Functional role of estrogen metabolism in target cells: review and perspectives](#). *Carcinogenesis* 19, 1–27.